

## ·综述·

# 血脑屏障与阿尔茨海默病病变的研究进展

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**摘要** 阿尔茨海默病(AD)是导致痴呆的重要原因,进行性认知、记忆及行动能力障碍是其临床特点。研究发现,超过40%的AD患者脑内存在血脑屏障(BBB)通透性增高现象,血管内皮细胞、周细胞损伤甚至死亡是其主要原因;同时血管内皮细胞、周细胞损伤又与痴呆的发生密切相关。本文综述总结AD与BBB的内在联系及作用机制的研究进展,以期为AD的治疗提供新的治疗方向。

**关键词** 阿尔茨海默病;血脑屏障;血管内皮细胞;周细胞

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血脑屏障(blood-brain barrier, BBB)是分隔大脑组织与外周环境的功能单位,由血管内皮细胞通过紧密连接蛋白(ZO-1、occludin、claudin-5等)、黏附连结蛋白相互紧密连接构成基本框架,周细胞及星形胶质细胞终足覆盖包裹内皮细胞所构成,进而限制外周大分子蛋白、细胞毒性物质、外周免疫细胞等进入神经中枢,维持中枢神经系统内环境稳定,维护脑细胞的正常功能<sup>[1]</sup>。研究证实,BBB与阿尔茨海默病(Alzheimer's disease, AD)的发生发展密切相关,同时也是防治AD的重要策略<sup>[2-5]</sup>。AD主要表现为进行性认知、记忆及行动能力障碍。神经元外淀粉样蛋白(Amyloid-β, Aβ)沉积、神经元内tau蛋白高度磷酸化形成神经纤维缠结、神经元丢失是AD的主要病理改变<sup>[6,7]</sup>。其病理机制复杂:早期主要是毒性蛋白Aβ激活炎症信号通路及Ca<sup>2+</sup>信号通路诱发神经元凋亡及神经突触丢失;疾病后期Aβ诱发神经元内tau蛋白高度磷酸化,致使其与微管蛋白结合降低,损伤神经元细胞结构,最终导致患者认知、记忆等能力障碍诱发痴呆<sup>[8-10]</sup>。血管内皮细胞及周细胞损伤是BBB破裂、血管通透性增加的主要原因,血管通透性增加致使外周有害物质进入脑内激活小胶质细胞,活化的小胶质细胞释放大量促炎因子如:白细胞介素(interleukin, IL)-1β、IL-6、肿瘤坏死因子(tumor necrosis factor, TNF)-α等,进而诱发神经元凋亡<sup>[11]</sup>;同时BBB破裂又能减少Aβ的外周清除,增加外周Aβ重新进入脑,增加脑内Aβ含量,损伤神经元,诱发痴呆<sup>[12]</sup>。因此,恢复BBB结构完整,降低血管通透性,增加Aβ清除,改善脑微环境,是治疗AD的新靶点。

## 1 BBB破裂增加脑内Aβ蓄积

研究发现,脑内生成的Aβ通过与BBB上载脂蛋白E(apolipoprotein E, APOE)结合而被转运到脑血管内皮细胞表面,进而与脂蛋白受体相关蛋白-1(lipoprotein receptor-related protein-1, LRP-1)结合,并在ATP结合盒式转运蛋白P-g glycoprotein(P-gP)

的协助下进行Aβ的外周清除,将Aβ跨BBB转运出脑<sup>[13,14]</sup>。同时,脑血管内皮细胞表达的晚期糖基化终产物(receptor for advanced glycation end products, RAGE)可与外周游离Aβ结合将其重新摄取入脑内<sup>[15]</sup>。当BBB上LRP-1含量降低,RAGE含量增高,将会导致Aβ外周清除受阻,重新入脑增多,加剧其脑内蓄积<sup>[16]</sup>。Anushruti等<sup>[17]</sup>发现慢性脑缺血时,表皮生长因子/heparin-binding EGF-like growth factor(HB-EGF)通过激活低氧诱导因子-1α(hypoxia-inducible factor-1alpha, HIF-1α)刺激基质金属蛋白酶9(matrix metalloproteinase-9, MMP-9)的表达,进而损伤脑血管内皮细胞,导致内皮细胞表面LRP-1含量降低,RAGE含量增高,Aβ重吸收增强,外周清除受阻,最终诱发神经元损伤,导致小鼠认知及记忆功能下降。

Abhay等<sup>[18]</sup>敲除AD转基因模型小鼠APP<sup>w<sup>o</sup>/0</sup>的血小板衍生生长因子受体β(platelet-derived growth factor β, pdgfrβ)基因复制周细胞缺失AD模型,结果发现小鼠脑内Aβ<sub>40</sub>、Aβ<sub>42</sub>水平较单纯的APP<sup>w<sup>o</sup>/0</sup>小鼠高,同时脑内血管淀粉样病变增多,海马及皮质淀粉样斑块数量也增加。进一步研究发现,周细胞缺失使Aβ半衰期延长,Aβ清除发生障碍;同时周细胞缺失还促进tau蛋白高度磷酸化,降低其与微管蛋白亲和力,导致神经元功能障碍。血管内皮细胞、周细胞都是BBB的细胞组成,两者的损伤缺失都将导致BBB结构破裂,导致Aβ外周清除受阻,发生脑内蓄积。同时,RAGE水平增高还将导致Aβ重新进入脑内,加重Aβ脑内蓄积。

## 2 BBB破裂诱发神经元死亡

脑组织通过BBB将脑细胞与外周分离开,一旦BBB结构破裂,屏障功能受损,必将导致外周毒性蛋白进入脑内,进而诱发神经元损伤。研究发现<sup>[19]</sup>,各种血管危险因素,如吸烟、高血压、糖尿病及各种脑血管病变均可导致周细胞、血管内皮细胞损伤、丢失,BBB通透性增高,外周纤溶酶、凝血酶、纤维

蛋白等进入脑实质,损伤神经元;此外,血浆白蛋白进入脑实质将导致血浆胶体渗透压改变,诱发颅内水肿,进而压迫神经血管,导致神经元低灌注发生,引发神经元的损伤。周细胞内表达平滑肌肌动蛋白- $\alpha$ (smooth muscle  $\alpha$ -actin,  $\alpha$ -SMA),还参与脑血流量的调节<sup>[20]</sup>。研究证实,周细胞缺失将导致脑血流量的减少,直接导致神经元能量代谢障碍、代谢废物不能及时清除,诱发神经元的死亡<sup>[21,22]</sup>。

### 3 A $\beta$ 导致 BBB 破裂的内在机制

对 APP/PS1 双转基因 AD 模型小鼠及 SAMP8 快速衰老 AD 模型小鼠的研究均发现,小鼠脑内血管内皮细胞损伤,BBB 通透性明显增高<sup>[23,24]</sup>,细胞实验发现血管内皮损伤与 A $\beta$ 密切相关。Yoonjin 等<sup>[25]</sup>利用微流体技术 3D 离体培养 ReN Cells(一种能够表达 APP/PSEN1 基因,进而合成 A $\beta$  的脑血管内皮细胞)发现 A $\beta$  水平增高,导致血管内皮细胞活性降低,细胞膜上紧密连接蛋白 claudin-1 及 claudin-5 蛋白水平降低,进一步检测发现 MMP-2 及活性氧水平增高,MMP-2 能够降解紧密连接蛋白,活性氧能诱发内皮细胞氧化损伤,导致 BBB 通透性增高;而减少 A $\beta$  生成后,BBB 通透性降低。Elvis 等<sup>[26]</sup>离体培养小鼠血管内皮细胞 rBMVEC 复制 BBB 模型,观察 A $\beta$  类似物质 A $\beta$ <sub>25-35</sub> 对 BBB 的影响,结果表明,细胞内活性氧水平增高,细胞活性降低,BBB 通透性增高。可见,在 AD 病变过程中,A $\beta$  能够诱发血管内皮细胞损伤,导致 BBB 破裂。

Matthew 等<sup>[27]</sup>对携带载脂蛋白 E4 (apolipoprotein E-4, APOE4) 的 AD 患者尸检发现,患者脑内周细胞数量明显较不携带 APOE4 的 AD 患者减少;对 APOE4 转基因小鼠研究发现,小鼠脑内 LRP-1 水平降低,导致 LRP-1 依赖的促炎信号通路亲环素 A(cyclophilin A, CypA)/MMP-9 被激活,进而诱发周细胞损伤。此外,Nina 等<sup>[28]</sup>对 AD 患者尸检发现,患者脑内周细胞数量减少与 A $\beta$ <sub>40</sub> 水平密切相关;研究证实纤维状 A $\beta$ <sub>40</sub> 处理周细胞后,细胞内半胱氨酸天冬氨酸蛋白酶 caspase3/7 活性增强,周细胞凋亡增加。在 AD 发展中,A $\beta$  能诱发血管内皮细胞、周细胞损伤导致 BBB 破裂,同时 AD 高致病基因 APOE4 亦能通过 LRP-1/CypA/MMP-9 信号通路诱发周细胞损伤。

### 4 血管内皮细胞、周细胞移植,修复 BBB 结构及功能可改善 AD 痴呆症状

Zhang 等<sup>[29]</sup>将血管内皮祖细胞植入 APP/PS1 转基因 AD 模型小鼠海马内,结果显示海马区毛细血管数量增多,紧密连接蛋白 ZO-1、occludin、claudin-5 表达增高,BBB 通透性降低;同时淀粉样斑块数量减少,A $\beta$  诱导的凋亡因子 Bax、caspase-3 活性降低,抗凋亡因子 bcl-2 表达增加,神经元凋亡减少;小鼠空间学习和记忆能力得到显著改善。提示改善脑微血管内皮细胞损伤可以有效治疗 AD。

同样的治疗效果在周细胞移植治疗 AD 中得到了验证。Masaya 等<sup>[30]</sup>将 C3H/10T1/2 小鼠间充质干细胞诱导分化为周细胞并注射入 APP/PS1 小鼠右半脑内,探究周细胞对 AD 的治疗作用。3 周后检测发现,右脑较左脑血液循环明显改善,同时脑

内 A $\beta$ <sub>40</sub>、A $\beta$ <sub>42</sub> 水平明显降低,海马区淀粉样斑块数量也显著降低,其内在机制是周细胞通过其表面的 LRP-1 增加了对 A $\beta$  的摄取,并通过其内在的 A $\beta$  降解酶降解 A $\beta$ ,减少脑内 A $\beta$  积淀。这提示,脑微血管内皮细胞与周细胞修复能够有效恢复 BBB 细胞及分子结构;恢复 BBB 屏障与转运功能改善认知及记忆损伤,是治疗 AD 的有效策略。

### 5 问题与展望

BBB 是脑实质与外周联系的桥梁,血管内皮细胞、周细胞是 BBB 重要的细胞组成,一旦二者发生损伤,将导致 BBB 功能障碍,进而诱发多种神经疾病,如 AD。同时,AD 也能导致血管内皮细胞、周细胞损伤,导致 BBB 功能障碍,加剧 AD 的病理改变。因此,修复 BBB 结构,恢复其功能具有延缓 AD 进展的作用。但如何促进内皮细胞、周细胞修复损伤 BBB,降低 AD 对 BBB 的损伤还需更深入的研究。

### 参考文献

- [1] Daneman R, Prat A. The blood-brain barrier[J]. Cold Spring Harb Perspect Biol, 2015, 7: a020412.
- [2] Yamazaki Y, Kanekiyo T. Blood-Brain Barrier Dysfunction and the Pathogenesis of Alzheimer's Disease[J]. Int J Mol Sci, 2017, 18: 1965.
- [3] Montagne A, Zhao Z, Zlokovic BV. Alzheimer's disease: A matter of blood-brain barrier dysfunction[J]. J Exp Med, 2017, 214: 3151-3169.
- [4] Cai Z, Qiao PF, Wan CQ, et al. Role of Blood-Brain Barrier in Alzheimer's Disease[J]. J Alzheimers Dis, 2018, 63: 1223-1234.
- [5] Zenaro E, Piacentino G, Constantin G. The blood-brain barrier in Alzheimer's disease[J]. Neurobiol Dis, 2017, 107: 41-56.
- [6] Schöll M, Ossenkoppele R, Strandberg O, et al. Distinct 18F-AV-1451 tau PET retentionpatterns in early- and late-onset Alzheimer's disease[J]. Brain, 2017, 140: 2286-2294.
- [7] Stocker H, Nabers A, Perna L, et al. Prediction of Alzheimer's disease diagnosis within 14 years through A $\beta$  misfolding in blood plasma compared to APOE4 status, and other risk factors[J]. Alzheimers Dement, 2020, 16: 283-291.
- [8] Tu S, Okamoto S, Lipton SA, et al. Oligomeric A $\beta$ -induced synaptic dysfunction in Alzheimer's disease[J]. Mol Neurodegener, 2014, 9: 48.
- [9] Jian M, Kwan JS, Bunting M, et al. Adiponectin suppresses amyloid- $\beta$  oligomer (A $\beta$  O)-induced inflammatory response of microglia via AdipoR1-AMPK-NF- $\kappa$ B signaling pathway[J]. J Neuroinflammation, 2019, 16: 110.
- [10] Ising C, Venegas C, Zhang S, et al. NLRP3 inflammasome activation drives tau pathology[J]. Nature, 2019, 575: 669-673.
- [11] Bhaskar K, Maphis N, Xu G, et al. Microglial derived tumor necrosis factor- $\alpha$  drives Alzheimer's disease-related neuronal cell cycle events[J]. Neurobiol Dis, 2014, 62: 273-285.
- [12] Löffler T, Flunkert S, Temmel M, et al. Decreased Plasma A $\beta$  in Hyperlipidemic APP/PS1 Transgenic Mice Is Associated with BBB Dysfunction[J]. Front Neurosci, 2016, 10: 232.
- [13] Storck SE, Hartz AMS, Bernard J, et al. The concerted amyloid-beta clearance of LRP1 and ABCB1/P-gp across the blood-brain barrier is linked by PICALM[J]. Brain Behav Immun, 2018, 73: 21-33.
- [14] Erdő F, Krajesi P. Age-Related Functional and Expressional Changes in Efflux Pathways at the Blood-Brain Barrier[J]. Front Aging Neurosci, 2019, 11: 196.
- [15] Cai Z, Liu N, Wang C, Qin B, et al. Role of RAGE in Alzheimer's Disease[J]. Cell Mol Neurobiol, 2016, 36: 483-495.
- [16] Shang J, Yamashita T, Tian F, et al. Chronic cerebral hypoperfusion alters amyloid- $\beta$  transport related proteins in the cortical blood vessels of Alzheimer's disease model mouse[J]. Brain Res, 2019, 15: 1723.
- [17] Ashok A, Rai NK, Raza W, et al. Chronic cerebral hypoperfusion-induced impairment of A $\beta$  clearance requires

- HB-EGF-dependent sequential activation of HIF1  $\alpha$  and MMP9[J]. *Neurobiol Dis*, 2016, 95: 179-193.
- [18] Sagare AP, Bell RD, Zhao Z, et al. Pericyte loss influences Alzheimer-like neurodegeneration in mice[J]. *Nat Commun*, 2013, 4: 2932.
- [19] Winkler EA, Sagare AP, Zlokovic BV. The pericyte: a forgotten cell type with important implications for Alzheimer's disease[J]. *Brain Pathol*, 2014, 24: 371-386.
- [20] Attwell D, Mishra A, Hall CN, et al. What is a pericyte[J]? *Cereb Blood Flow Metab*, 2016, 36: 451-455.
- [21] Fernández-Klett F, Priller J. Diverse functions of pericytes in cerebral blood flow regulation and ischemia[J]. *Cereb Blood Flow Metab*, 2015, 35: 883-887.
- [22] Hall CN, Reynell C, Gesslein B, et al. Capillary pericytes regulate cerebral blood flow in health and disease[J]. *Nature*, 2014, 508: 55-60.
- [23] Cao Y, Xu H, Zhu Y, et al. ADAMTS13 maintains cerebrovascular integrity to ameliorate Alzheimer-like pathology[J]. *PLoS Biol*, 2019, 17: e3000313.
- [24] PelegríC, Canudas AM, del Valle J, et al. Increased permeability of blood-brain barrier on the hippocampus of a murine model of senescence [J]. *Mech Ageing Dev*, 2007, 128: 522-528.
- [25] hin Y, Choi SH, Kim E, et al. Blood-Brain Barrier Dysfunction in a 3D In Vitro Model of Alzheimer's Disease[J]. *Adv Sci (Weinh)*, 2019, 6: 1900962.
- [26] Cuevas E, Rosas-Hernandez H, Burks SM, et al. Amyloid Beta 25-35 induces blood-brain barrier disruption in vitro[J]. *Metab Brain Dis*, 2019, 34: 1365-1374.
- [27] Halliday MR, Rege SV, Ma Q, et al. Accelerated pericyte degeneration and blood-brain barrier breakdown in apolipoprotein E4 carriers with Alzheimer's disease. [J]. *Cereb Blood Flow Metab*, 2016, 36: 216-227.
- [28] Schultz N, Brännström K, Byman E, et al. Amyloid-beta 1-40 is associated with alterations in NG2+ pericyte population ex vivo and in vitro[J]. *Aging Cell*, 2018, 17: e12728.
- [29] Zhang S, Zhi Y, Li F, et al. Transplantation of in vitro cultured endothelial progenitor cells repairs the blood-brain barrier and improves cognitive function of APP/PS1 transgenic ADmice[J]. *Neurol Sci*, 2018, 387: 6-15.
- [30] Tachibana M, Yamazaki Y, Liu CC, et al. Pericyte implantation in the brain enhances cerebral blood flow and reduces amyloid- $\beta$  pathology in amyloid model mice[J]. *Exp Neurol*, 2018, 300: 13-21.

(本文编辑:唐颖馨)

(上接第447页)

- [4] Powers WJ, Rabinstein AA, Ackerson T, et al. 2018 Guidelines for the Early Management of Patients With Acute Ischemic Stroke: A Guideline for Healthcare Professionals From the American Heart Association/American Stroke Association[J]. *Stroke*, 2018, 49: e46-e110.
- [5] 陆小波, 肖国栋, 肖章红, 等. 神经介入取栓术治疗脑梗死的疗效研究[J]. 神经损伤与功能重建, 2019, 14: 523-524.
- [6] Horsch AD, Dankbaar JW, van der Graaf Y, et al. Relation between reperfusion and hemorrhagic transformation in acute ischemic stroke[J]. *Neuroradiology*, 2015, 57: 1219-1225.
- [7] Balian NR, Alonso CB, Zurru MC, et al. Predictores clínicos de transformación hemorrágica en accidente cerebrovascular isquémico no lacunar [Clinical predictors of hemorrhagic transformation in non lacunar ischemic stroke][J]. *Medicina (B Aires)*, 2017, 77: 100-104.
- [8] Neuberger U, Möhlenbruch MA, Herweh C, et al. Classification of Bleeding Events: Comparison of ECASS III (European Cooperative Acute Stroke Study) and the New Heidelberg Bleeding Classification[J]. *Stroke*, 2017, 48: 1983-1985.
- [9] Yaghi S, Willey JZ, Cucchiara B, et al. Treatment and Outcome of Hemorrhagic Transformation After Intravenous Alteplase in Acute Ischemic Stroke: A Scientific Statement for Healthcare Professionals From the American Heart Association/American Stroke Association[J]. *Stroke*, 2017, 48: e343-e361.
- [10] van Kranendonk KR, Treurniet KM, Boers AMM the MR CLEAN investigators, et al. Hemorrhagic transformation is associated with poor functional outcome in patients with acute ischemic stroke due to a large vessel occlusion[J]. *J Neurointerv Surg*, 2019, 11: 464-468.
- [11] Barber PA, Demchuk AM, Zhang J, et al. Validity and reliability of a quantitative computed tomography score in predicting outcome of hyperacute stroke before thrombolytic therapy. ASPECTS Study Group. Alberta Stroke Programme Early CT Score[J]. *Lancet*, 2000, 355: 1670-1674.
- [12] Schröder J, Thomalla G. A Critical Review of Alberta Stroke Program Early CT Score for Evaluation of Acute Stroke Imaging[J]. *Front Neurol*, 2017, 7: 245.
- [13] Wardlaw JM, von Kummer R, Farrall AJ, et al. A large web-based observer reliability study of early ischaemic signs on computed tomography. The Acute Cerebral CT Evaluation of Stroke Study (ACCESS)[J]. *PLoS One*, 2010, 5: e15757.
- [14] MacCallum C, Churilov L, Mitchell P, et al. Low Alberta Stroke Program Early CT score (ASPECTS) associated with malignant middle cerebral artery infarction[J]. *Cerebrovasc Dis*, 2014, 38: 39-45.
- [15] Liu L, Wu B, Zhao J, et al. Computed Tomography Perfusion Alberta Stroke Program Early Computed Tomography Score Is Associated with Hemorrhagic Transformation after Acute Cardioembolic Stroke[J]. *Front Neurol*, 2017, 8: 591.
- [16] Asuzu D, Nystrom K, Amin H, et al. Comparison of 8 scores for predicting symptomatic intracerebral hemorrhage after IV thrombolysis[J]. *Neurocrit Care*, 2015, 22: 229-233.
- [17] Kalinin MN, Khasanova DR, Ibatullin MM. The hemorrhagic transformation index score: a prediction tool in middle cerebral artery ischemic stroke[J]. *BMC Neurol*, 2017, 17: 177.
- [18] Albers GW, Marks MP, Kemp S, et al. Thrombectomy for Stroke at 6 to 16 Hours with Selection by Perfusion Imaging[J]. *N Engl J Med*, 2018, 378: 708-718.
- [19] Song L, Lyu C, Shen G, et al. Application of FLAIR Vascular Hyperintensity-DWI Mismatch in Ischemic Stroke Depending on Semi-Quantitative DWI-Alberta Stroke Program Early CT Score[J]. *Front Neurol*, 2019, 10: 994.
- [20] Zimmerman RD, Maldjian JA, Brun NC, et al. Radiologic estimation of hematoma volume in intracerebral hemorrhage trial by CT scan[J]. *AJR Am J Neuroradiol*, 2006, 27: 666-670.
- [21] Liu ZW, Jiang Y, Wang R, et al. Combined application of multi-parameter semiquantitative Alberta stroke program early CT score to assess infarction severity in acute ischemic stroke[J]. *Zhonghua Yi Xue Za Zhi*, 2018, 98: 1697-1702.
- [22] Guberina N, Dietrich U, Radbruch A, et al. Detection of early infarction signs with machine learning-based diagnosis by means of the Alberta Stroke Program Early CT score (ASPECTS) in the clinical routine [J]. *Neuroradiology*, 2018, 60: 889-901.
- [23] Murray NM, Unberath M, Hager GD, et al. Artificial intelligence to diagnose ischemic stroke and identify large vessel occlusions: a systematic review[J]. *J Neurointerv Surg*, 2020, 12: 156-164.
- [24] Albers GW, Wald MJ, Mlynash M, et al. Automated Calculation of Alberta Stroke Program Early CT Score: Validation in Patients With Large Hemispheric Infarct[J]. *Stroke*, 2019, 50: 3277-3279.
- [25] Kuang H, Najim M, Chakraborty D, et al. Automated ASPECTS on Noncontrast CT Scans in Patients with Acute Ischemic Stroke Using Machine Learning[J]. *AJR Am J Neuroradiol*, 2019, 40: 33-38.

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